

IMMUNOMODULATORY ACTIVITY OF LEAVES OF *RUMEX VESICARIUS* LINN. AND *SYMPLOCOSRACEMOSA* ROXB.

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ABSTRACT: Objective: In order to find out how *Rumex vesicarius* Linn. and *Symplocos racemosa* Roxb. leaves affect the immune system, we used the following methods: The *Rumex Vesicarius* Linn. ethyl acetate and ethanolic extract, as well as the *Symplocos racemosa* Roxb. ethanolic and N-hexane extract, were given orally to rats at doses of 200 mg/kg/day and 400 mg/kg/day, respectively, based on their body weight. Evaluations of the carbon clearance test and the delayed type hypersensitivity test were used to measure the immunomodulatory action. The results demonstrated a substantial increase in phagocytic activity and DTH response in rats when the extracts of *Rumex vesicarius* Linn. and *Symplocos racemosa* Roxb. were administered orally at an experimental dosage ($P < 0.0001$). Acute toxicity and preliminary phytochemical screening are also part of the research. Results suggest that both humoral and cell-mediated immunity are significantly impacted by the immunomodulatory effects of the *Rumex vesicarius* Linn. and *Symplocos racemosa* Roxb. extracts, according to the study's conclusion.

Keywords: *Rumex vesicarius* Linn., *Symplocos racemosa* Roxb., Carbon Clearance Test, Delayed Type Hypersensitivity test

INTRODUCTION: The immune system is a remarkably versatile defense system that has evolved to protect animals from invading pathogenic microorganisms to eliminate the disease. It is able to generate a multifarious variety of cells and molecules capable of specifically recognizing and eliminating variety of foreign invaders¹. It is now being recognized that modulation of immunological response could provide an alternative to conventional chemotherapy for a variety of diseased conditions of impaired immuneresponsiveness or when a selective immunosuppression has to be induced in situations like autoimmune disorders and organ transplantation.

The modulation of the immune response by using Ayurvedic herbal medications as a possible therapeutic measure has now become a subject of scientific investigation. One of the therapeutic strategies in Ayurvedic medicines is to enhance the body's overall natural resistance to the disease causing agent rather than directly neutralizing the agent itself². Immunomodulation is the process of modifying an immune response in a positive (immunostimulation) or negative manner (immunosuppressant) by administration of a drug or compound³.

There are many plants which are used as immunomodulators. *Heterostemma tanjorensis* shows immunostimulant activity against Azathioprine administered rats⁴. The methanolic root extract of *Withania somnifera* shows immunostimulatory activities in dexamethasone induced immunocompromised mice and in vitro model⁵. The Methanolic leaf extract of *Moringa oleifera* shows an immunostimulatory effect on

both the cell-mediated and humoral immune systems in the Wistar albino rats⁶. The methanolic leaf extracts of *Cameroonian* medicinal plants possess immunomodulatory activity⁷. The ethanolic extract of *Sonerilatinneveli* showed a stimulatory effect on both humoral and cellular immune functions in animal models⁸. *Caesalpinia sappan* shows the nonspecific immunomodulatory effect on murine peritoneal macrophages⁹. The aqueous leaf extract of *Ocimum basilicum* Linn. is a potent immunostimulant, stimulating specific and nonspecific immune mechanisms. The immunostimulatory activity of *O. basilicum* is due to flavonoids (quercetin), alkaloids, tannins, saponin glycosides and phenolic compounds¹⁰.

The methanolic extract of *Trigonella foenum-graecum* whole plant has shown significant immunostimulatory activity in various models of Drug-induced myelosuppression¹¹. The flavonoids isolated from freshly harvested leaves of *Prosopis spicigera*, *Mimusops elengi* and *Terminalia arjuna* showed dose dependent immunosuppressive activity¹². The crude terpenoids extract from the leaves of *Emblica officinalis*, *Ficus racemosa* and *Strychnos nux-vomica* on human whole blood stimulated with the hepatitis B vaccine possess immunosuppressive activity¹³.

The aqueous extract of *Leucas aspera* was evaluated in cyclophosphamide-induced immunosuppressive mice and it shows prominent immunostimulatory effect¹⁴. The leaves of *Calotropis gigantea*, *C. rotang* and *A. integrifolia* have an immunosuppressive activity of the variable doses of crude saponin (0.625–2.5 mg) on lymphocytes, monocytes and granulocytes¹⁵.

Triptolide, the active component of *Tripterygium wilfordii* can be used alone or in combination with existing therapeutic modalities as novel treatments for autoimmune disorders, cancers, and for immunosuppression¹⁶. The ethanolic extracts from leaves of *Rhaphidophora korthalsii* stimulate immune cell proliferation, peripheral blood NK cell population¹⁷. Cyclotides, ribosomally synthesized plant peptides have growth-inhibiting effects on primary cells of the human immune system¹⁸. The isogarcinol, active compound from *Garcinia mangostana* L. inhibits Calcineurin unique protein phosphatase, plays an important role in immunoregulation in a dose-dependent manner¹⁹. Natural therapies help to regulate the immune system's aggressive behavior without suppressing necessary defenses²⁰.

For this study we have selected two plants, *Rumex vesicarius* Linn. and *Symplocos racemosa* Roxb.

Rumex vesicarius Linn. (Chooka) belongs to perennial herb to the family Polygonaceae. The plant is an erect usually with a long taproot. Traditionally the plant is used as stomachic, Diuretic, used for the disorders of the lymphatic and glandular system, for bronchitis, asthma, constipation, dyspepsia and the diseases of the liver. Plant leaves are rich in ascorbic acid, citric acid and tartaric acid, it also contains glycoside, alkaloid, flavonoids, tannins and phenolic compounds^{21,22}.

Symplocos racemosa Roxb. (Lodhra) belongs to the family Symplocaceae, is a small evergreen tree up to 6 m tall. In traditional system it is mainly used as a cardiotonic, antipyretic, antihelminthic and laxative properties. It is beneficial in bilious fever, urinary discharge; pharmacologically it is used as an antimicrobial, antidiarrhoeal, spasmogenic and heart depressant. The plant mainly contains monomethyl pelargonidin glucosides, loturidine also contain oxalic acid, phytosterol, ellagic acids and oleonic acid^{23,24,25}.

MATERIALS AND METHODS:

Plant Material: The fresh leaves of *Rumex vesicarius* and *Symplocos racemosa* Roxb. used in this study, collected at the flowering stage (Month: August - November) from the local area of Sangli and Satara, Maharashtra state, India respectively and authenticated by Botanical Survey of India, Pune, Maharashtra. (BSI/WRC/Iden./2015 dated 4-12-2015)

Extraction: The leaves were separated from fresh stems and dried under shade at room temperature until it becomes completely dry. After drying leaves were subjected to size reduction. The shade-dried coarsely powdered leaves (500g) were subjected to Soxhlet extraction.

A) *Rumex vesicarius* Linn. leaves were subjected to Soxhlet extraction with 95% ethanol and ethyl

acetate to obtain ethanolic and ethyl acetate extract respectively.

B) *Symplocos racemosa* Roxb. leaves were subjected to Soxhlet extraction with 95% ethanol and N-hexane to obtain ethanolic and N-hexane extract respectively. The extracts obtained were subjected to the Rotary flash evaporator to remove excess of solvent and dried extracts were stored in a cool place in tight pack container for further use.

Animals: All the experiments were carried out using male albino rats of wistar strain. Weight around 150-200 gm. The animals are free to access of food and water, and they were housed in a natural (12 h each) light-dark cycle. The animals were acclimatized for at least 5 days to the laboratory conditions before the experiment. The experimental protocol was approved by the institutional animal ethics committee (IAEC/ABCP/09/2016-17) and the care of laboratory animal was taken as per the guidelines of CPCSEA, the ministry of forests and environment government of India.

Preliminary Phytochemical Screening: All the extracts were subjected to preliminary phytochemical screening using the method described by Kokate, Trease and Evans for the detection of various plants constituents. Test were carried out for the presence or absence of Phytoconstituents like glycosides, flavonoids, saponins, alkaloids, carbohydrates, sterols, phenolic compound and reducing compounds^{26,27,28}.

Drugs and Chemicals: All the drugs and Chemical were of analytical grade while the other drugs were procured from - Levamisole (Johnson and Johnson Ltd.), Cyclophosphamide (Biochem pharmaceutical industries Ltd.), Colloidal carbon (Indian ink, Camel India Pvt. Ltd.).

Selection of doses: Acute Toxicity

studies were performed according to the organization for economic cooperation and development (OECD) guideline (425). For acute toxicity study there was no mortality and physical/behavioral changes showed after administration of all the extract over 14 days at the dose of 2000 and 5000 mg/kg to a different group of rat weight around 150 - 200 gm. The experiments were performed after the experimental protocols had been approved by the institutional animal ethical committee.

Pharmacological Screening: The immuno-modulatory activity is carried out by using Carbon clearance test. (Test for Phagocytosis) and Delayed-Type Hypersensitivity Reaction.

Carbon Clearance Test: (Test for Phagocytosis): Procedure:

- In this test, animals were divided into 11 groups comprising 6 animals in each.

TABLE 1: TREATMENT GROUPS OF CARBON CLEARANCE TEST

Groups	Treatment	Dose and route of administration
Group I	Vehicle	10ml/kg P.O.
Group II	Standard drug (Cyclophosphamide)	50mg/kg P.O.
Group III	Standard drug (Levamisole)	2.5mg/kg P.O.
Group IV	Ethyl acetate extract of leaves of <i>Rumex vesicarius</i> L. (EARV-400)	400mg/kg P.O.
Group V	Ethyl acetate extract of leaves of <i>Rumex vesicarius</i> L. (EARV-200)	200mg/kg P.O.
Group VI	Ethanolic extract of leaves of <i>Rumex vesicarius</i> L. (ERV-400)	400mg/kg P.O.
Group VII	Ethanolic extract of leaves of <i>Rumex vesicarius</i> L. (ERV-200)	200mg/kg P.O.
Group VIII	Ethanolic extract of leaves of <i>Symplocos racemosa</i> Roxb. (ESR-400)	400mg/kg P.O.
Group IX	Ethanolic extract of leaves of <i>Symplocos racemosa</i> Roxb. (ESR-200)	200mg/kg P.O.
Group X	n-hexane extract of leaves of <i>Symplocos racemosa</i> Roxb. (NSR-400)	400mg/kg P.O.
Group XI	n-hexane extract of leaves of <i>Symplocos racemosa</i> Roxb. (NSR-200)	200mg/kg P.O.

- Carbon ink suspension was injected via tail vein to each rat 48 hours after the five day treatment
- Blood samples (25 µl) were then withdrawn from the retro-orbital plexus with mild ether anesthesia at 5 and 15 min after injection of colloidal carbon ink lysed in 0.1% sodium carbonate solution (3ml).
- The optical density was measured spectrophotometrically at 660 nm.
- The phagocytic activity was calculated using the following formula^{29,30}.

$$K = \frac{\text{LogOD}_1 - \text{LogOD}_2}{t_2 - t_1}$$

Where ODI and OD2 are the optical density at time t_1 and t_2 , respectively.

Preparation of Carbon Ink Suspension: Camlin ink was diluted eight times with saline and used for carbon clearance test in a dose of 10 µl/gm body weight of rat³¹.

Statistical Analysis: The result was expressed as mean value \pm SEM. The variation in a set of data has been estimated by performing one-way analysis of variation (ANOVA). Individual comparison of group mean value were done using Dunnett's test. The P value < 0.05, were considered statistically significant.

Delayed Type of Hypersensitivity Reaction: Procedure:

- In this test, animals were divided into 11 groups comprising 6 animals in each.

TABLE 2: TREATMENT GROUPS OF DELAYED TYPE HYPERSENSITIVITY

Groups	Treatment	Dose and route of administration
Group I	Vehicle	10ml/kg P.O.
Group II	Standard drug (Cyclophosphamide)	50mg/kg P.O.
Group III	Standard drug (Levamisole)	2.5mg/kg P.O.
Group IV	Ethyl acetate extract of leaves of <i>Rumex vesicarius</i> L. (EARV-400)	400mg/kg P.O.
Group V	Ethyl acetate extract of leaves of <i>Rumex vesicarius</i> L. (EARV-200)	200mg/kg P.O.
Group VI	Ethanol extract of leaves of <i>Rumex vesicarius</i> L. (ERV-400)	400mg/kg P.O.
Group VII	Ethanol extract of leaves of <i>Rumex vesicarius</i> L. (ERV-200)	200mg/kg P.O.
Group VIII	Ethanol extract of leaves of <i>Symplocos racemosa</i> Roxb. (ESR-400)	400mg/kg P.O.
Group IX	Ethanol extract of leaves of <i>Symplocos racemosa</i> Roxb. (ESR-200)	200mg/kg P.O.
Group X	N-hexane extract of leaves of <i>Symplocos racemosa</i> Roxb. (NSR-400)	400mg/kg P.O.
Group XI	N-hexane extract of leaves of <i>Symplocos racemosa</i> Roxb. (NSR-200)	200mg/kg P.O.

- Immunized Rat with 0.1ml of 20% SRBCS in normal saline intraperitoneally on 14th day of the study. On day 21st, animals from all groups get challenge with 0.03 ml of 1% SRBCs in subplantar region of the right hind paw. Footpad reaction was assessed after 24 hrs *i.e.* on the 22nd day. Increase in footpad edema was measured with the help of vernier caliper²⁹.

Antigenic Material:

Preparation of Sheep RBCs: Sheep blood was collected in sterile Alsever's solution in 1:1 proportion, Alsever's solution (freshly prepared) blood was kept in the refrigerator and processed for the preparation of SRBCs batch, by centrifugation at 2000 rpm for 10 min and washing with physiological saline 4-5 times and then suspending into buffered saline for further use³¹.

Composition of Alsever's Solution:

TABLE 3: COMPOSITION OF ALSEVER'S SOLUTION

Chemicals	Quantity (g/L)
Sodium Chloride	4.2
Sodium Citrate	8.0
Citric acid anhydrous	0.55
Glucose	20.5
Distilled water q.s.	1000 ml

Statistical Analysis: The result was expressed as mean value \pm SEM. The variation in a set of data has been estimated by performing one-way analysis of variation (ANOVA). Individual comparison of group mean value were done using Dunnett's test. The P value < 0.05 , were considered statistically significant.

RESULT:

Acute Oral Toxicity Study: Acute oral toxicity was carried out by the up-down method. It is found that all extract (EARV, ERV, ESR and NSR) were safe at limit dose 4000 mg/kg and 2000 mg/kg, with no mortality and physical/behavioral changes. 1/10th of this dose *i.e.* 400 mg/kg and 200 mg/kg were used in the subsequent study.

Preliminary Phytochemical Study: The presence of various phytoconstituents of the extract was detected by phytochemical screening. The EARV found to contain Alkaloids, Flavonoids, Tannins, Sterols, Carbohydrate and Vitamin C. ERV contains Alkaloids, Flavonoids, Carbohydrate and Vitamin C. ESR found to contain Cardiac glycoside, Flavonoids, Alkaloids, Tannins and Carbohydrate. NSR contains cardiac glycoside, alkaloids and steroids.

Carbon Clearance Test: Effect of EARV, ERV, ESR, and NSR on the phagocytic activity by the carbon clearance test is shown in **Table 4**. The phagocytic activity of the reticuloendothelial system is generally measured by the rate of removal of carbon particles from the bloodstream. In carbon clearance test EARV, ERV and ESR,

TABLE 4: RESULT OF CARBON CLEARANCE TEST

treated all groups exhibited significantly high phagocytic index ($P < 0.0001$) when compared with control group. While NSR treated group showed a small increase in their phagocytic index when compared with control group. This indicates stimulation of the reticuloendothelial system.

S.no.	Groups	Treatments	Dose and route of administration	Phagocytic index (Mean \pm SEM)
1	I	Control	10ml/kg (P.O.)	0.0312 \pm 0.0005
2	II	Standard (Cyclophosphamide)	50mg/kg (P.O.)	0.0206 \pm 0.0005****
3	III	Standard (Levamisole)	2.5mg/kg (P.O.)	0.0573 \pm 0.0003****
4	IV	Ethyl acetate extract of leaves of <i>Rumex vesicarius</i> L. (EARV-400)	400mg/kg (P.O.)	0.0530 \pm 0.0004****
5	V	Ethyl acetate extract of leaves of <i>Rumex vesicarius</i> L. (EARV-200)	200 mg/kg (P.O.)	0.0492 \pm 0.0004****
6	VI	Ethanol extract of leaves of <i>Rumex vesicarius</i> L. (ERV-400)	400mg/kg (P.O.)	0.04566 \pm 0.0005****
7	VII	Ethanol extract of leaves of <i>Rumex vesicarius</i> L. (ERV-200)	200mg/kg (P.O.)	0.0414 \pm 0.0003****
8	VIII	Ethanol extract of leaves of <i>Symplocos racemosa</i> Roxb. (ESR-400)	400mg/kg (P.O.)	0.0391 \pm 0.0003****
9	IX	Ethanol extract of leaves of <i>Symplocos racemosa</i> Roxb. (ESR-200)	200mg/kg (P.O.)	0.0363 \pm 0.0004****
10	X	N-hexane extract of leaves of <i>Symplocos racemosa</i> Roxb. (NSR-400)	400mg/kg (P.O.)	0.0350 \pm 0.0002****
11	XI	N-hexane extract of leaves of <i>Symplocos racemosa</i> Roxb. (NSR-200)	200mg/kg (P.O.)	0.0327 \pm 0.0002

Values are expressed as (Mean \pm SEM). N=6****P<0.0001. Statistically significant when compared with control group by ANOVA followed by Dunnett's test.

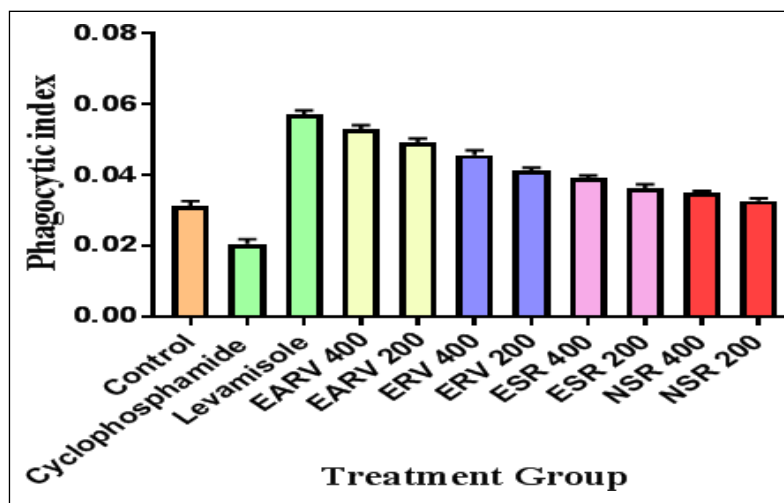


FIG.1: GRAPHICAL REPRESENTATION OF CARBON CLEARANCE TEST

Delayed Type of Hypersensitivity Reaction: Effect of EARV, ERV, ESR and NSR on the cell-mediated immune response by DTH induce footpad edema is shown in **Table 5**. All treated groups EARV, ERV and ESR showed the significantly ($p < 0.0001$) potentiating DTH response in terms of increase in the mean difference of paw edema when compared with control group. It indicates activation of the cellular immune system. While NSR treated group showed a small increase in footpad oedema when compared with control group. Cyclophosphamide treated group showed a significant decrease in the mean difference of paw edema when compared with control group.

TABLE 5: RESULT OF DTH

S. no.	Groups	Treatments	Dose and route of administration	Mean Difference in Paw edema (Mean \pm SEM)
1	I	Control	10ml/kg(P.O.)	1.508 \pm 0.0316
2	II	Standard(Cyclophosphamide)	50mg/kg(P.O.)	0.59 \pm 0.0513****
3	III	Standard(Levamisole)	2.5mg/kg(P.O.)	4.56 \pm 0.0594****
4	IV	Ethylacetate extract of leaves of <i>Rumex vesicarius</i> L.(EARV-400)	400mg/kg(P.O.)	4.20 \pm 0.0545****
5	V	Ethylacetate extract of leaves of <i>Rumex vesicarius</i> L.(EARV-200)	200mg/kg(P.O.)	3.66 \pm 0.123****
6	VI	Ethanol extract of leaves of <i>Rumex vesicarius</i> L.(ERV-400)	400mg/kg(P.O.)	3.22 \pm 0.0519****
7	VII	Ethanol extract of leaves of <i>Rumex vesicarius</i> L.(ERV-200)	200mg/kg(P.O.)	2.51 \pm 0.0586****
8	VIII	Ethanol extract of leaves of <i>Symplocos racemosa</i> Roxb.(ESR-400)	400mg/kg(P.O.)	2.29 \pm 0.0152****
9	IX	Ethanol extract of leaves of <i>Symplocos racemosa</i> Roxb.(ESR-200)	200mg/kg(P.O.)	2.08 \pm 0.0345****
10	X	N-hexane extract of leaves of <i>Symplocos racemosa</i> Roxb (NSR-400)	400mg/kg(P.O.)	1.90 \pm 0.0085****
11	XI	N-hexane extract of leaves of <i>Symplocos racemosa</i> Roxb.(NSR-200)	200mg/kg(P.O.)	1.74 \pm 0.02986*

Values are expressed as (Mean \pm SEM). n=6****P<0.0001. Statistically significant when compared with control group by ANOVA followed by Dunnett's test.

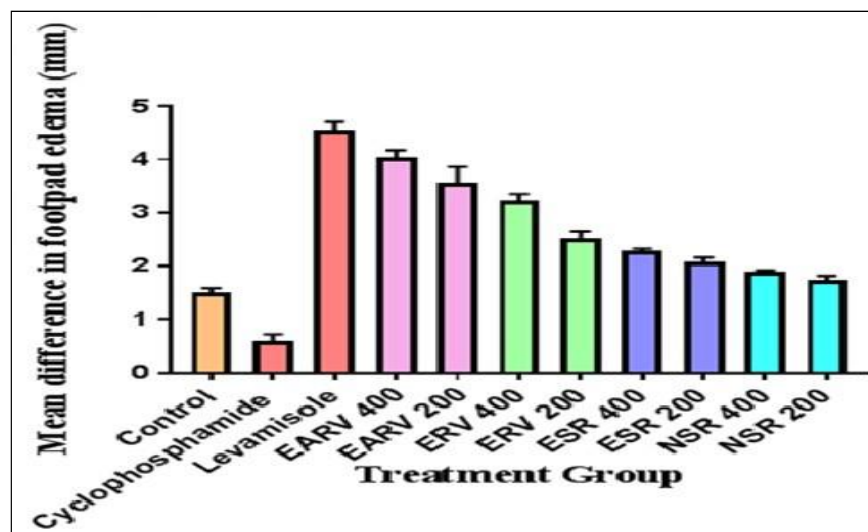


FIG. 2: GRAPHICAL REPRESENTATION OF DTH

DISCUSSION: The immune system is a remarkably versatile system that has evolved to defend itself against a vast range of harmful agents. It is able to generate an enormous variety of cell and molecules capable of specifically recognizing and eliminating a variety of foreign invaders. Immunomodulators are a natural or synthetic substance that helps to regulate or normalize the immune system. Immunomodulators correct immune systems that are out of balance. And immunomodulation is a process which can alter the immune system specifically immunostimulation and immunosuppressant.

The present study was designed to explore the immunomodulatory activity of *Rumex vesicarius* Linn. and *Symplocos racemosa* Roxb. In this study Carbon Clearance Test and Delayed Type Hypersensitivity test were selected for evaluation of immunomodulatory activity of *Rumex vesicarius* Linn. and *Symplocos racemosa* Roxb.

According to Smrithi Tripathi *et al.*, the Phagocytic activity of reticuloendothelial system was assayed by carbon clearance test phagocytic index was calculated as the rate of carbon elimination of reticuloendothelial system.

In the present study after oral administration of all the extract (EARV, ERV, ESR and NSR) at an experimental dose (400 mg/kg and 200 mg/kg) showed a significant increase in the phagocytic index ($P < 0.0001$) when compared with control group. Increase in phagocytic activity indicates that there was stimulation of reticuloendothelial system.

According to N. L. Dashputre *et al.*, Cell-mediated immunity involves the interaction of effector mechanism carried out by T lymphocytes and their products (lymphokines). DTH requires specific recognition of antigen by activated T lymphocytes, which subsequently proliferate and release cytokines. These in turn increase vessel permeability cause vasodilatation, macrophage accumulation, and activation promoting phagocytic activity and increased concentration of lysozyme for more effective killing. The delay in the onset of the response reflects the time required for the cytokine to induce the recruitment and activation of macrophages. Therefore increase in DTH response after oral administration of all the extract (EARV, ERV, ESR and NSR) at experimental dose (400 mg/kg and 200 mg/kg) showed a significant increase in foot pad edema ($P < 0.0001$) when compared with control. It indicates stimulation of the cell-mediated immunity.

The present study revealed that ethyl acetate extract of *Rumex vesicarius* Linn. showed highest immunomodulatory activity.

In this study, the overall order of immuno-modulatory activity was established as EARV > ERV > ESR > NSR.

CONCLUSION: Both humoral and cell-mediated immunity were found to be significantly impacted by the present study's ethanolic and methyl acetate extracts of *Rumex vesicarius* Linn. and *Symplocos racemosa* Roxb. leaves, respectively. This was attributed to the following mechanisms:

- The activation of T-cells, which mediated the DTH response;
- The activation of the reticuloendothelial system.
- Raise the efficiency of the macrophage system's monocytes.

Additionally, when comparing the immunomodulatory activity of the four extracts (EARV, ERV, ESR, and NSR), the ethyl acetate extract of *Rumex vesicarius* Linn. leaves (EARV) exhibited the highest activity at the experimental dosage. As a whole, EARV is more immunomodulatory than ERV, ESR is more immunomodulatory than NSR, and so on. Its precise immunomodulatory effect mechanism may be determined by a thorough examination.

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